

# Review Article

# Stimulatory effect of hormonal seed priming in plant tolerance to resist abiotic stress

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## Abstract

Plants exposing abiotic stresses such as drought, salinity, heat, cold, and heavy metals that induce complex responses ultimately result in reduced growth as well as crop yield. Phytohormones are widely known for their regulatory functions in controlling plant growth and development. They also act as significant chemical messengers, enabling plants to survive when subjected to a variety of stressors. Nowadays various strategies are employed that can withstand these emergences. In recent years, seed priming has been an indispensable method to induce tolerance against various stresses. The seed priming process is a physiological method that involves hydration for enhancement of seed germination, early seedling growth, and yield under stressed and non-stressed conditions. The seedlings emerging from primed seeds showed early and uniform germination. Moreover, the overall growth of plant is enhanced due to the seed-priming treatments with phytohormone which have become a significant strategy for reducing the impacts of abiotic stress. Therefore, this review analyses the potentiality of priming with several phytohormones to mitigate the negative impacts of abiotic stresses, for improving crop productivity.

## 1. Introduction

There are some of the major kinds of stresses like heat, drought, cold, and salt stress that crops usually face under adverse weather or soil conditions. Disturbance equilibrium which produces changes in in physiological parameters, and due to stress plant's chemical and physiological changes occur is called stress [1]. In most plants, stress causes a variety of biochemical, physiological, and metabolic changes [2], which may result in oxidative stress and affect plant metabolism, performance, and thereby yield [3]. often interrelated, Abiotic stresses are either individually or in combination; they cause morphological, physiological, biochemical, and molecular changes that affect plant growth and development and ultimately yield. In the present era

of global climate change, abiotic stresses are becoming more prevalent. The increasing threat of climate change is already having a substantial impact on agricultural production worldwide causing significant unpredictable loss in agriculture [4] and threat to global food security [5].

Plants are subjected to a variety of abiotic stress such as salinity, drought, high temperature, low temperature, etc. which reduces germination rate and seedling growth with significant variations from crop to crop [6]. Salinity has an adverse effect on seed germination and seedling growth of several crops either by creating an osmotic potential in the rhizosphere of the plant that inhibits the absorption of water or creates toxic effects to the roots and whole



crop because of Na<sup>+</sup> and Cl<sup>-</sup> [7, 8]. Drought is one of the most important environmental factors limiting plant growth and productivity. With the increase of drought severity, the drought severity increased, the germination rate linearly decreased in unprimed cotton seeds [9]. Low-temperature conditions decreased plant growth rate because of inhibition of photosynthesis and increasing photo-oxidative injury of the photosystems [10]. Photo-oxidative damage caused lipid peroxidation and degradation of chlorophyll and carotene [110]. Plants exert many physiological and biochemical changes under lowtemperature conditions that make them survive under these conditions [10]. Heat stress is often defined as the rise in temperature beyond a threshold level for a period of time sufficient to cause irreversible damage to plant growth and development. The extent to which it occurs in specific climatic zones depends on the probability and period of high temperatures occurring during the day and/or the night [11]. Thus, abiotic stress causes many physiological and biochemical changes in the seedlings, which include the generation of reactive oxygen species [ROS], leading to membrane damage and cell leakage and destruction of photosynthetic components [12].

Various methodologies were adapted from time to time to achieve tolerance against stresses. These include conventional breeding methods such as selection and hybridization and modern methods such as mutation breeding, genetic engineering, etc. [12]. Attempts were also made to produce transgenic plants which can withstand various kinds of stresses [12]. But these methods are time-consuming and demand skills and involve legal and ethical issues. The alternative solution would be more acceptable if it is simple, cost-effective, and can be adopted by the farmers without any complication, and at the same time, it should be effective in manifesting the tolerance.

Seed priming is one such farmer's friendly technique recommended by many researchers for better crop stand establishment and growth even under adverse conditions. It is a simple, safe, economic, and effective approach for enhancement of seed germination, early seedling growth and yield under stressed and nonstressed conditions [13]. In plant defence, priming is defined as a physiological process by which a plant prepares to respond to imminent abiotic stress more

#### quickly or aggressively.

The priming process induces the rate of seed germination and is associated with the initiation of germination-related processes [14] and repair processes [15] and increases various free radicalscavenging enzymes, such as catalase, and peroxidase [16]. Several seed priming methods were successfully used in agriculture for seed conditioning to accelerate the germination rate and improve the seedling uniformity [17, 18]. Moreover, seed priming helps many crops to neutralize the adverse effects of abiotic stress [19]. The various approaches of seed priming are hydro priming, osmopriming, chemical priming, hormonal priming, biological priming, redox priming, solid matrix priming, etc. [111]. Among these techniques, seed priming with phytohormones (hormonal priming) has emerged as a promising strategy in modern stress management as it protects plants against various abiotic stresses by increasing the level of antioxidant enzyme activity, decreasing oxidative damage, and enhancing plant growth. Priming for enhanced resistance to abiotic stress is operating via various pathways involved in different metabolic processes. It is known that seed priming can activate these signalling pathways in the early stages of growth and result in faster plant defence responses. Therefore, the purpose of this review is to summarise the understanding of the regulation mechanism against abiotic stresses through hormonal priming to mitigate the losses occurred in crop production in future.

#### 2. Materials and methods

Relevant literature on hormonal Seed Priming was composed for plant growth and yield attributing activities released up to January 2023. The literature has been searched on the hormonal priming activity of the different phytohormones on different crops. keywords were: The main abiotic tolerant, phytohormones, plant growth, priming, yield etc. GoogleScholar®, ResearchGate®, Web of Science®, PubMed, SciFindern and Scopus® were used as electronic search tools for articles with the several definite keywords. We have reviewed only the manuscripts which are relevant to this article.

## 3. Results and discussion

#### 3.1 Phytohormones

Plant hormones are known as phytohormones or

plant growth regulators (PGRs). These are chemical molecules produced by plants and have important roles in regulating plant growth and development (Fig. 1). Phytohormones function as important chemical messengers and modulate many cellular processes in plants and can coordinate different signalling pathways during exposure to abiotic stresses [20, 21]. Auxins (IAAs), cytokinins (CKs), gibberellins [GAs], abscisic acid (ABA), salicylic acid and ethylene (ET) are well known [SA], phytohormones, essential for plant growth and development [22, 23].

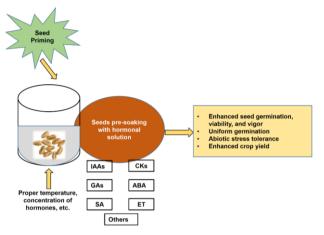


Figure 1. Schematic model showing possible effects of seed priming with phytohormones [24]

#### 3.2 Hormonal priming

Seed priming with hormone solutions is referred to as hormonal priming, and hormonal seed priming plays an important role in seed metabolism [24]. Seeds are pre-soaked with an optimal concentration of phytohormone, which enhances germination, seedling growth and yield by increasing nutrient uptake through enhanced physiological activities and root production [25, 26]. Commonly used plant growth regulators in seed priming are IAAs, CKs, GAs, ABA, SA, and ET.

#### 3.3 Auxins

The role of auxin in plant development is well known; however, its possible function in response to various stresses is poorly understood (Fig. 2). Several studies demonstrate a novel role of auxin signalling and transport in plant tolerance to abiotic stress [27]. Seed IAAs enhances cell priming with division, photosynthetic activities, and translocation of carbohydrates, which results in lateral root initiation, flowering, and good stand establishment [28]. Seed priming with IAAs (1 ppm) enhanced the seedling

establishment of Bouteloua gracilis [29], and in wheat grass (Agropyron elongates), seeds priming with IAAs at 50 ppm improved tolerance to drought stress by enhancing antioxidant enzyme activities such as catalase [CAT], superoxide dismutase [SOD], and peroxidase [30]. Auxin positively modulates root biomass and branching, which might improve water uptake efficiency as well as partly participates in the positive regulation of drought stress resistance through the regulation of root architecture [31]. According to [31], auxin positively regulated the activities of four enzymatic antioxidants (superoxide dismutase, catalase, peroxidase, glutathione reductase) under drought stress conditions, thus conferring effective ROS (reactive oxygen species) detoxification to improve drought stress resistance.

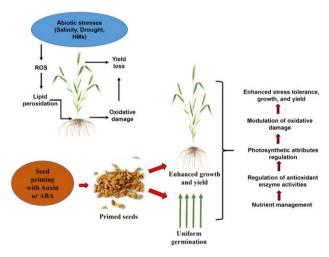


Figure 2. Proposed possible mechanisms used by auxin-and abscisic acid (ABA) priming and their roles on the germination, growth, and development of plants under different stresses [24].

Under salinity stress, wheat seeds priming with IAAs (100, 150 and 200 mg L<sup>-1</sup>) regulated hormonal homeostasis, which enhanced the CO<sub>2</sub> assimilation rate and ultimately resulted in increased grain yield [32]. SOS pathway (which maintains ion homeostasis under salt stress) modulates root response by regulating PIN2 protein and auxin asymmetric distribution [33]. Also, seed priming with IAAs improved the germination and growth of different species, such as rice (*Oryza sativa*] and pigeon pea (*Cajanus cajan*), under arsenic or cadmium (Cd) stress [34].

High and low day and night temperature (suppose 24-35 °C day temperature and 5-10 °C night temperature) was found to reduce fruit set, pollen grain viability,

Plant	Stresses	Responses of Plant	References
Soybean (Glycine max)	Drought	Improved drought tolerance in soybean plants	[55]
Pigeon pea (Cajanus cajan)	Salt	Prevented the damage caused by the apparatus involved in protein synthesis	[57]
	Cadmium	Tolerance to the effects of Cd stress	[34]
Basil (Ocimum basilicum)	Drought	Reduced negative effects of drought stress	[58]
Wheat (Triticum aestivum)	Salt	Decreased ABA concentration, increased IAAs concentration, and enhancement of salt tolerance	[59]
	Salt	Improved photosynthetic rate, water use efficiency and stomatal conductance, decreased Na <sup>+</sup> and Cl <sup>-</sup> level, increased K <sup>+</sup> level	[60]
	Salt	Decreased electrolyte leakage and conferred salt tolerance	[61]
	Salt	Increased tissue N content and nitrate reductase activity	[62]
	Salt	Induced reduction in inorganic ion accumulation and increasing membranes stability and K <sup>+</sup> /Na <sup>+</sup> ratio, enhanced chlorophyll formation and soluble sugar accumulation	[63]
	Salt	Alleviated salt stress by enhanced ethylene production	[64]

Table 1. Seed-priming with cytokinin adopted for developing abiotic stress tolerance in plants.

and IAA levels in tomato [35]. However, application of auxin completely reversed male sterility in barley and *Arabidopsis* [36]. The content of auxin was not affected by proline, but the expression of auxin carriers was reduced and in the overexpression lines of PDH, in which proline content was reduced, the expression of auxin carrier genes was induced [114].

#### 3.4 Cytokinin

The exogenous application of CKs can mitigate the abiotic stresses on crop plants, which ultimately results in increased growth, development, and yield [52]. Likewise, supplementation of CKs also reduces salinity stress in plants [52], and it increases starch accumulation in salt-stressed rice plants [53] (Table 1). It has been reported that wheat seeds priming with kinetin (100 mg L<sup>-1</sup>, 150 mg L<sup>-1</sup>, and 200 mg L<sup>-1</sup>) enhanced germination and tolerance against salt by decreasing ABA and increasing IAAs concentrations [54]. Likewise, Mangena [55] reported that soybean seed priming with CKs (Benzyl adenine; 4.87 mg L-1) increased soybean root biomass, flowering, and fruiting under drought stress. Priming of aged groundnut (Arachis hypogaea L.) seeds with CKs (150 ppm) enhanced germination and seedling indices by enhancing antioxidant enzyme activities and decreasing oxidative damage [56]. Seed priming with CKs or a combination of CKs and other plant hormones has resulted in the mitigation of abiotic stresses in various plant species.

#### 3.5 Gibberallin

Different abiotic stresses, such as salinity, drought, chilling, heat, and heavy metals, inhibit proper nutrient uptake and photosynthesis, which ultimately results in stunted plant growth [65]. The exogenous application of gibberallin can mitigate abiotic stresses and enhance plant growth and development (Table 2). Exogenous application of gibberallin improved the growth of wheat (Triticum aestivum) plants and mitigated drought induced oxidative damage by maintaining relative water content, balancing the antioxidant mechanism system, and conserving the chlorophyll concentration [66]. Foliar application of GA3 @50ppm to tomato (Solanum lycopersicum) plants increased relative leaf water content, stomatal density, and chlorophyll content by mitigating salinity stress [67]. Besides, GA3 was stimulated in plant growth and vield leaf of lettuce (Lactuca sativa) by enhancing biomass accumulation, leaf expansion, stomatal conductance, water use efficiency, and nitrogen use efficiency [68].

#### 3.6 Abscisic acid

ABA is one of the major plant hormones and is also

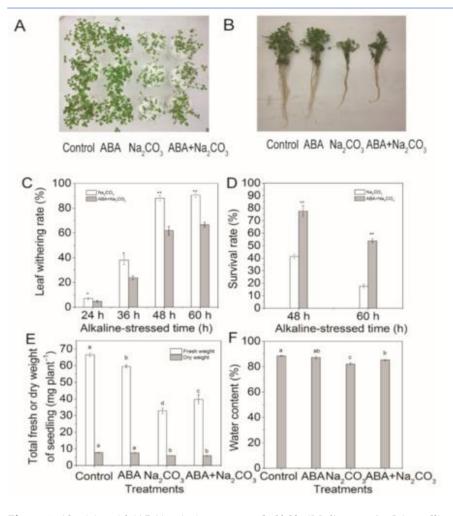
Table 2. Seed-priming with gibberellin and respo	onse of plant species.
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Plants	Stresses	Responses of Plant	References
Marigold and Sweet fennel	Salt	Increased dry matter and enhanced tolerance to salinity by enhancing antioxidant enzyme activities	[13]
Pigeon pea (Cajanus cajan)	Cadmium	Increased germination speed index and germination percentage and tolerance to Cd stress	[34]
Milk Thistle (Silybum marianum)	Salt	Increased $\alpha$ -amylase activity and alleviated salt stress effects	[37]
Chickpea ( <i>Cicer arietinum</i> )	Drought	Increased relative water content, seed protein, and reduced electrolyte leakage	[38]
Wheat (Triticum aestivum)	Salt	Promoted better salinity tolerance	[39]
Sorghum (Sorghum bicolor)	Drought	Increased CAT and APX activities	[40]
Corn (Zea mays)	Salt	Increased tissue water content	[41]
Maize ( <i>Zea mays</i> ), Pea ( <i>Pisum sativum</i> ), Grass pea ( <i>Lathyrus sativus</i> )	Salt	Alleviated salt stress effects	[42]
Rice (Oryza sativa)	Flood	Increased $\alpha$ -Amylase activity, sucrose, glucose, and fructose content in seeds.	[43]
Alfalfa (Medicago sativa)	Salt	Induced enzymatic activities (SOD, CAT, GPX, APX, GR) and decreased lipid peroxidation, and reduced membrane damage of alfalfa.	[44]
Sponge gourd ( <i>Luffa aegyptiaca</i> )	Salt	Prevented the adverse effect of salinity	[45]
Soybean ( <i>Glycine max</i> )	Saline- alkali	Increased activities of the antioxidant defense system, photosynthetic pigment contents, better membrane integrity	[46]
Maize (Zea mays)	Salt	Reduced negative effect of salt stress	[47]
Sweet sorghum (Sorghum bicolor)	Salt	Enhanced water absorption and improved salinity tolerance	[48]
Maize (Zea mays)	Drought	Increased chlorophyll content and enhance drought tolerance	[49]
Okra (Abelmoschus esculentus)	Salt	Increased water content of the okra seedlings	[50]
Triticale	Salt	Reduced Na <sup>+</sup> accumulation and increased K <sup>+</sup> uptake	[51]

known as a stress hormone. It plays a vital role in mediating plant responses to various abiotic stresses, such as salt, heat, and drought [69] (Fig. 3). ABA not only plays a role in abiotic stress mitigation but also plays a significant role in plant growth and development [70]. Rice seeds primed with ABA exhibited enhanced seedling growth and yield in saline soil by balancing nutrient uptake [71]. Likewise, priming rice seeds with ABA reduced alkaline stress by enhancing antioxidant enzyme activities and the activity of stress tolerance-related genes in the roots of rice seedlings [72]. It has been reported that phytohormones are effective in the mitigation of heavy metal stress [23]. ABA biosynthetic gene expressions are induced by heavy metal stresses, which results in increased levels of endogenous ABA [73]. Priming *Arabidopsis* seeds with amino-butyric acid enhanced drought tolerance by accumulation of ABA and the closing of stomata [74]. The regulation of proline metabolism is dependent on ABA accumulation [116], whereas other responses occur independently of ABA, and that ABA alone cannot duplicate drought-induced proline accumulation [117]. They proposed that GAs inhibited flowering and ABA promoted flowering in litchi [118].

#### 3.7 Salicylic acid

Salicylic acid (SA) is a phenolic compound involved in the regulation of growth and development of plants, and their responses to biotic and abiotic stress



**Figure 3.** Abscisic acid (ABA) priming protected alfalfa (*Medicago sativa* L.) seedlings from wilting and death under alkaline conditions. Eighteen-day-old alfalfa seedlings were root-drenched with 10  $\mu$ M ABA or without 10  $\mu$ M ABA (Control) for 16 h and then exposed to alkaline stress (15 mM Na<sub>2</sub>CO<sub>3</sub>). (**A**, **B**) Photographs of seedling growth were taken after 60 h of alkaline treatment, (**C**) leaf withering (%) was recorded at 24 h, 36 h, 48 h, and 60 h, (**D**) survival rate of alfalfa seedlings was determined after 48 h and 60 h of alkaline treatment, (**E**) total fresh or dry weight of seedlings and (**F**) water content were measured after 60 h of alkaline treatment. Values are the mean ± standard error, n = 3. Asterisks denote a significant difference compared with control plants (\* *p* < 0.05, \*\* *p* < 0.01) based on Student's *t*-test. Different letters above the columns indicate significant differences (*p* < 0.05) at each time point based on Duncan's test [112].

factors [75] (Table 3; Fig. 4). It is involved in the regulation of important plant physiological processes such as photosynthesis, nitrogen metabolism, proline metabolism, production of glycine betaine, antioxidant defense system, and plant-water relations under stress conditions and thereby protects plants against abiotic stresses [75]. SA has been shown to improve plant tolerance to major abiotic stresses such as metal [76], salinity [77], drought [78], and heat stress [79]. The exogenous application of salicylic acid enhanced maize (Zea mays) productivity under low temperature stress, as well as the germination and growth parameters of garden cress (Lepidium sativum) seedlings under salinity stress [80], and mitigated drought stress and enhanced the vegetative growth of

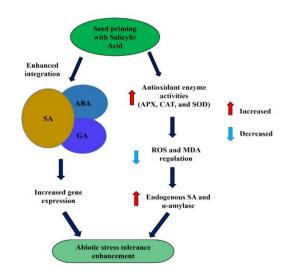


Figure 4. Mechanisms of SA priming for abiotic stress tolerance enhancement [24].

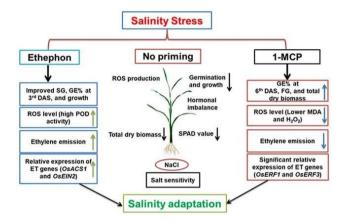
Table 3. Seed priming with salicylic acid and response of plant species.

Crops	Stresses	Responses of Plants	References
Rice (Oryza sativa)	Chromium	Increased chlorophyll content and proper nutrient uptake	[85]
	Water deficit	Decreased water stress	[86]
	Chilling	Enhanced antioxidant enzyme activities, detoxified ROS	[87]
	Salinity	Improved Na <sup>+</sup> /K <sup>+</sup> and maintaining membrane integrity	[88]
Safflower ( <i>Carthamus tinctorius</i> )	Drought	Enhanced antioxidant enzyme activities and reduced oxidative damage	[89]
Maize (Zea mays)	Chilling	Increased $\alpha$ -amylase and antioxidant enzyme activities and endogenous SA content	[90]
	Chilling	Enhanced enzymatic antioxidant activities, high tissue water content	[91]
	Lead	Increased glycine betaine and nitric oxide content and regulation of gene expression	[92]
	Chromium and UV-B	Reduced the accumulation of chromium and ROS	[93]
Wheat ( <i>Triticum aestivum</i> )	Salinity	Decreased the electrolyte leakage	[94]
	Drought	Balanced nutrient uptake	[95]
	Osmotic	Resistance to osmotic stress	[96]
	Salinity	Higher contents of photosynthetic pigments, soluble sugar, and protein	[97]
	Boron toxicity	Increased photosynthetic pigments	[98]
	Cadmium	Modulates nutrient relations and photosynthetic attributes	[99]
Smooth vetch (Vicia dasycarpa)	Water deficit	Higher accumulation of proline and glycine betaine	[100]
Okra (Abelmoschus esculentus)	Chilling	Enhanced antioxidant enzyme activities and membrane integrity	[101]
Sorghum (Sorghum bicolor)	Drought	Improved antioxidant defense system	[102]
Tomato (Solanum lycopersicum)	Salinity	Decreased salinity stress	[103]
	Heat	Increased lycopene content	[104]
Pumpkin	Salinity	Protein contents and nitrate reductase were increased	[105]
Faba bean ( <i>Vicia faba</i> )	Salinity	Higher osmotic solute content, carotenoids, and antioxidant enzyme activity	[106]

drought stress and enhanced the vegetative growth of safflower (*Carthamus tinctorius*) [81]. Priming with salicylic acid at 100 mg L<sup>-1</sup> enhanced emergence and produced early seedling growth in cucumber (*Cucumis sativus*) [82] and increased germination and productivity of *Vicia faba* [83] and sesame (*Sesamum indicum*) [84].

#### 3.8 Ethylene

Ethylene is regarded as a stress-responsive hormone besides its role in regulation of plant growth and development [107] (Fig. 5). The hydrocarbon ethylene is an important plant hormone and it is widely used for ripening fruits [108]. It was found that exogenous application of ethylene with sufficient sulphur level counteracted the cadmium-induced photosynthetic and growth inhibition in mustard plants [109]. Free proline content and ethylene production increase in cold-acclimated winter rapeseed seedlings under pretreatment with glutamine and especially with proline [115]. Free proline is involved in the response to cold stress, and its level may be an indicator of coldhardening and freezing tolerance, but the role of ethylene in the regulation of cold tolerance remains not quite clear [115].



**Figure 5.** Ethylene response of salt stressed rice seedlings following Ethephon and 1-methylcyclopropene seed priming. SG= speed of germination, GE= germination energy percentage, 3rd and 6th DAS= 3rd and 6th day after sowing, and FG= final germination [113].

## 4. Conclusions

Seed priming with phytohormones has emerged as a promising strategy in modern stress management as it protects plants against various abiotic stresses by increasing level of antioxidant enzyme activity, decreasing oxidative damage, and enhancing plant growth. Thus, seed priming with phytohormones improves the tolerance of crop plants to abiotic stress, and this technique can be utilized to maintain sustainable crop production in drought-, saline-, and flood-prone areas of the world. Seed priming with phytohormones not only improves the tolerance to abiotic stresses but also ensures harmonized germination by breaking the dormancy and enhancing viability. This review provides insight into the role of seed priming with phytohormones in mitigating the effects of abiotic stress on seed germination and plant growth. Seed priming with phytohormones has emerged as an effective seed treating tool for many crops, but treating conditions and methods differ from crop to crop, and seed priming with phytohormones has still limitations. For instance, prolonged seed treatment with hormonal solution during priming may cause the loss of seed tolerance to desiccation, which reduces seed viability.

## **Authors' contributions**

All authors equally contributed, read and approved the final manuscript.

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## Availability of data and materials

All data will be made available on request according to the journal policy.

## **Conflicts of interest**

There is no potential conflict of interest to describe.

## References

- Gaspar, T.; Franck, T.; Bisbis, B.; Kevers, C.; Jouve, L.; Hausman, J.F.; Dommes, J. Concepts in plant stress physiology. Application to plant tissue cultures. Plant Growth Regul. 2002, 37(3), 263-285.
- Xiong, L.; Zhu, J.K. Molecular and genetic aspects of plant responses to osmotic stress. Plant Cell Environ. 2002, 25, 131-139.
- Shafi, M.; Bakht, J.; Hassan, M.J.; Raziuddin, M.; Zhang, G. Effect of cadmium and salinity stresses on growth and antioxidant enzyme activities of wheat (*Triticum aestivum* L). Bull. Environ. Contam. Toxicol. 2009, 82, 772-776.
- Jakab, G.; Ton, J.; Flors, V.; Zimmerli, L.; Me'traux, J.P.; Mauch-Mani, B. Enhancing Arabidopsis salt and drought stress tolerance by chemical priming for its abscisic acid responses. Plant Physiol. 2005, 139, 267-274.
- Christensen, J.H.; Christensen, O.B. A summary of the PRUDENCE model projections of changes in European climate by the end of this century. Clim Change. 2007, 81, 7-30.
- Hamidi, H.; Safarnejad, A. Effect of drought stress on alfalfa cultivars (*Medicago sativa* L.) in germination stage. Am. Eurasian. J. Agric. Environ. Sci. 2010, 8(6), 705-709.
- Khajeh-Hosseini, M.; Powell, A.A.; Bimgham, I.J. The interaction between salinity stress and seed vigor during germination of soybean seeds. Seed Sci Technol. 2010, 31, 715-725.

- 8. Munns, J; Tester M. Mechanisms of salinity tolerance. Ann. Rev. Plant Biol. 2003, 59, 651-681.
- Soltani, A.; Gholipoor, M.; Zeinali, E. Seed reserve utilization and seedling growth of wheat as affected by drought and salinity. Environ. Exp. Bot. 2006, 55, 195-200.
- Xin, Z.; Browse, J. Cold comfort farm: the acclimation of plants to freezing temperatures. Plant Cell Environ. 2000, 23, 893-902.
- Wahid, A.; Gelani, S.; Ashraf, M.; Foolad, M. Heat tolerance in plants: an overview. Environ. Exp. Bot. 2007, 61,199-223.
- Jisha, K.C.; Puthur, J.T. Seed priming with BABA (βamino butyric acid): a cost-effective method of abiotic stress tolerance in *Vigna radiata* (L). Wilczek. Protoplasma. 2015, doi:10.1007/s00709-015-0804-7.
- Sedghi, M.; Nemati, A.; Esmailepour, B. Effect of seed priming on germination and seedling growth of two medicinal plants under salinity. Emir. J. Food. Agric. 2010, 22, 130-139.
- 14. Soeda, Y.; Konings, M.C.J.M.; Vorst, O.; Van Houwelingen, A.M.M.L.; Stoopen, G.M.; Maliepaard, C.A.; Kodde, J.; Bino, R.J.; S.P.C.G.; van der Geest, A.H.M. Gene expression programs during *Brassica oleracea* seed maturation, osmopriming, and germination are indicators of progression of the germination process and the stress tolerance level. Plant Physiol. 2005, 137, 354-368.
- 15. Sivritepe, H.O.; Dourado, A.M. The effect of priming treatments on the viability and accumulation of chromosomal damage in aged pea seeds. Ann. Bot. 1995, 75, 165-171.
- Gallardo, K.; Job, C.; Groot, S.P.C.; Puype, M.; Demol, H.; Vandekerckhove, J.; Job, D. Proteomic analysis of *Arabidopsis* seed germination and priming. Plant Physiol. 2001, 126, 835-848.
- Nouman, W.; Siddiqui, M.T.; Basra, S.M.A.; Afzal, I.; Rehman, H.U. Enhancement of emergence potential and stand establishment of *Moringa oleifera* Lam. by seed priming. Turk J Agric. 2012, 36, 227-235.
- Aghbolaghi, M.A.; Sedghi, M. The effect of osmo and hormone priming on germination and seed reserve utilization of millet seeds under drought stress. J. Stress Physiol. Biochem. 2014, 10(1), 214-221.
- 19. Ashraf, M.; Foolad, M.R. Pre-sowing seed treatment: a shotgun approach to improve germination, plant growth and crop yield under saline and non-saline conditions. Adv. Agron. 2005, 88, 223-271.
- Vob, U.; Bishopp, A.; Farcot, E.; Bennett, M.J. Modelling hormonal response and development. Trends Plant Sci. 2014, 19, 311-319.
- Kazan, K. Diverse roles of jasmonates and ethylene in abiotic stress tolerance. Trends Plant Sci. 2015, 20, 219-229.

- 22. Muhei, S.H. Seed priming with phytohormones to improve germination under dormant and abiotic stress conditions. Adv. Crop Sci. Technol. 2018, *6*, 403-409.
- Sytar, O.; Kumar, P.; Yadav, S.; Brestic, M.; Rastogi, A. Phytohormone priming: Regulator for heavy metal stress in plants. J. Plant Growth Reg. 2018, 38, 739-752.
- 24. Rhaman, M.S.; Rauf, F.; Tania, S.S.; Khatun, M. Seed priming methods: Application in field crops and future perspectives. Asian J. Res. Crop Sci. 2020, 5, 8-19.
- 25. Afzal, I.; Basra, S.M.A.; Ahmad, N.; Cheema, M.A.; Warriach, E.A.; Khaliq, A. Effect of priming and growth regulator treatment on emergence. Int. J. Agric. Biol. 2002, 4, 306.
- Akbari, G.; Sanavy, S.A.; Yousefzadeh, S. Effect of auxin and salt stress (NaCl) on seed germination of wheat cultivars (*Triticum aestivum* L.). Pak. J. Biol. Sci. 2007, 10, 2557-2561.
- 27. Krisnhamurthy, A.; Rathinasabapathi, B. Auxin and its transport play a role in plant tolerance to arsenite-induced oxidative stress in *Arabidopsis thaliana*. Plant, Cell and Environment. 2013, 36(10), 1838-1849.
- 28. MacDonald, H. Auxin perception and signal transduction. Physiol. Plant. 1997, 100, 423-430.
- 29. Roohi, R.; Jameson, D.A. The effect of hormone, dehulling and seedbed treatments on germination and adventitious root formation in blue grama. J. Range Manag. 1991, 44, 237 -241.
- 30. Eisvand, H.R.; Tavakkol-Afshari, R.; Sharifzadeh, F.; Maddah Arefi, H.; Hesamzadeh Hejazi, S.M. Effects of hormonal priming and drought stress on activity and isozyme profiles of antioxidant enzymes in deteriorated seed of tall wheatgrass (*Agropyron elongatum*). Seed Sci. Technol. 2010, 38, 280-297.
- Shi, H.; Chen, L.; Ye, T.; Liu, X., Ding, K. Chan, Z. Modulation of auxin content in Arabidopsis confers improved drought stress resistance. Plant Physiol. Biochem. 2014, 82, 209-217.
- 32. Iqbal, M.; Ashraf, M. Seed treatment with auxins modulates growth and ion partitioning in salt stressed wheat plants. J. Integr. Plant Biol. 2007, 49, 1045-1057.
- 33. Sun, F.; Zhang, W.; Hu, H.; Li, B.; Wang, Y.; Zhao, Y.; Li, K.; Liu, M.; Li, X. Salt modulates gravity signalling pathway to regulate growth direction of primary roots in arabidopsis. Plant Physiol. 2008, 146(1), 178-188.
- Sneideris, L.C.; Gavassi, M.A.; Campos, M.L.; Damico-Damiao, V.; Carvalho, R.F. Effects of hormonal priming on seed germination of pigeon pea under cadmium stress. Anais da Acad. Brasileira de Ciências 2015, 87, 1847-1852.
- El-Abd, S.O.; El-Beltagy, A. S.; Hall, M.A. Physiological studies on flowering and fruit set in tomatoes. Acta Horticul. 1986,190, 389-396.
- 36. Sakata, T.; Oshino, T.; Miura, S.; Tomabechi, M.; Tsunaga, Y.; Higashitani, N.; Miyazawa, Y.; Takahashi,

H.; Watanabe, M.; Higashitani, A. Auxins reverse plant male sterility caused by high temperatures. Agric. Sci. 2010, 107(19), 8569-8574.

- 37. Sedghi M.; Nemati, A.; Amanpour-Balaneji, B.; Gholipouri, A. Influence of different priming materials on germination and seedling establishment of milk thistle (*Silybum marianum*) under salinity stress. World Appl. Sci. J. 2010, 11604-609.
- Shariatmadari, M.H.; Parsa, M.; Nezami, A.; Kafi, M. Effects of hormonal priming with gibberellic acid on emergence, growth and yield of chickpea under drought stress. Biosci. Res. 2017, 14,34-41.
- Abido, W.A.E.; Allem, A.; Zsombik, L.; Attila, N. Effect of gibberellic acid on germination of six wheat cultivars under salinity stress levels. Asian J. Biol. Sci. 2019, 12, 51-60. doi: 10.3923/ajbs.2019.51.60.
- Sheykhbaglou, R.; Rahimzadeh, S.; Ansari, O.; Sedghi, M. The Effect of salicylic acid and gibberellin on seed reserve utilization, germination and enzyme activity of sorghum (*Sorghum bicolor* L) seeds under drought stress. J. Stress Physiol. Biochem. 2014, 10-5-13.
- Ghodrat, V.; Rousta, M.J. Effect of priming with gibberellic acid (GA<sub>3</sub>) on germination and growth of corn (*Zea mays* L.) under saline conditions. Int. J. Agric. Crop. Sci. 2012, 4, 883-885.
- Tsegay, B.A.; Andargie, M. Seed priming with gibberellic acid (GA<sub>3</sub>) alleviates salinity induced inhibition of germination and seedling growth of *Zea* mays L., Pisum sativum Var. abyssinicum A. Braun and Lathyrus sativus L. J. Crop Sci. Biotechnol. 2018, 21, 261-267. doi: 10.1007/s12892-018-0043-0.
- 43. Watanabe, H.; Honma, K.; Adachi, Y.; Fukuda, A. Effects of combinational treatment with ethephon and gibberellic acid on rice seedling growth and carbohydrate mobilization in seeds under flooded conditions. Plant Prod. Sci. 2018, 21, 380-386. doi: 10.1080/1343943X.2018.1520048.
- Younesi, O.; Moradi, A. Effect of priming of seeds of *Medicago sativa* 'Bami' with gibberellic acid on germination, seedlings growth and antioxidant enzymes activity under salinity stress. J. Hortic. Res. 2014, 22, 167–174. doi: 10.2478/johr-2014-0034.
- 45. Raheem, S.; Khan, J.; Gurmani, A.R.; Waqas, M.; Hamayun, M.; Khan A.L.; Kang, S.M.; Lee, I.J. Seed priming with gibberellic acid (GA<sub>3</sub>) in sponge gourd modulated high salinity stress. Pakhtunkhwa J. Life Sci. 2014, 2, 75-86.
- Dai, L.Y.; Zhu, H.D.; Yin, K.D.; Du, J.D.; Zhang, Y.X. Seed priming mitigates the effects of saline-alkali stress in soybean seedlings. Chil. J. Agric. Res. 2017, 77, 118-125. doi: 10.4067/S0718-58392017000200118.
- 47. Hamza, J.H.; Ali, M.K.M. Effect of seed soaking with GA<sub>3</sub> on emergence and seedling growth of corn under

salt stress. Iraqi J. Agric. Sci. 2017, 48, 560–566. doi: 10.36103/ijas. v48i3.377.

- Zhu, G.; An, L.; Jiao, X.; Chen, X.; Zhou, G.; McLaughlin, N. Effects of gibberellic acid on water uptake and germination of sweet sorghum seeds under salinity stress. Chil. J. Agric. Res. 2019, 79, 415–424. doi: 10.4067/S0718-58392019000300415.
- 49. Nada, H.S.; Hamza, J.H. Priming of maize seed with gibberellin [GA<sub>3</sub>] to tolerate drought stress. 2. Field emergence and its properties. Iraqi J. Des. Stud. 2019, *9*, 1-12.
- Yakoubi, F.; Babou, F.Z.; Belkhodja, M. Effects of gibberellic and abscisic acids on germination and seedling growth of okra (*Abelmoschus esculentus* L.) under salt stress. Pertanika J. Trop. Agric. Sc. 2019, 42, 847-860.
- Samad, R.; Karmokar, J.L. Effects of gibberellic acid and kn on seed germination and accumulation of Na<sup>+</sup> and K<sup>+</sup> in the seedlings of Triticale-I under salinity stress. Bangladesh J. Bot. 2012, 41, 123-129. doi: 10.3329/bjb. v41i2.13435.
- 52. Ha, S.; Vankova, R.; Yamaguchi-Shinozaki, K.; Shinozaki, K.; Tran, L.S. Cytokinins: Metabolism and function in plant adaptation to environmental stresses. Trends Plant Sci. 2012, *17*, 172-179.
- 53. Javid, M.G.; Sorooshzadeh, A.; Sanavy, S.A.M.M.; Allahdadi, I.; Moradi, F. Effects of the exogenous application of auxin and cytokinin on carbohydrate accumulation in grains of rice under salt stress. Plant Growth Reg. 2011, *65*, 305–313. doi: 10.1007/s10725-011-9602-1.
- Iqbal, M.; Ashraf, M.; Jamil A. Seed enhancement with cytokinins: Changes in growth and grain yield in salt stressed wheat plants. Plant Growth Reg. 2006, 50, 29-39. doi: 10.1007/s10725-006-9123-5.
- 55. Mangena, P. Effect of hormonal seed priming on germination, growth, yield and biomass accumulation in soybean grown under induced drought stress. Indian J. Agric. Res. 2020.
- 56. Sepehri, A.; Rouhi, H.R. Effect of cytokinin on morphological and physiological characteristics and antioxidant enzymes activity of aged groundnut (*Arachis hypogaea* L.) seeds under drought stress. Iran. J. Seed Sci. Technol. 2016, 5, 181-198.
- 57. Verma, J.; Srivastava, A.K. Physiological basis of salt stress resistance in pigeon pea (*Cajanus cajan* L.). II. Presowing seed soaking treatment in regulating early seedling metabolism during seed germination. Plant Physiol. Biochem. 1998, 25, 89-94.
- Bagheri, A.; Bagherifard, A.; Saborifard, H.; Ahmadi, M.M.; Safarpoor, M. Effects of drought, cytokinin and GA<sup>3</sup> on seedling growth of basil (*Ocimum basilicum*). Int. J. Adv. Biol. Biomed. Res. 2014, *2*, 489-493.

- Iqbal, M.; Ashraf, M. Presowing seed treatment with cytokinins and its effect on growth, photosynthetic rate, ionic levels and yield of two wheat cultivars differing in salt tolerance. J. Integr. Plant Biol. 2005, 47, 1315– 1325. doi: 10.1111/j.1744-7909.2005.00163. x.
- Afzal, I.; Basra, S.M.; Iqbal, A. The effects of seed soaking with plant growth regulators on seedling vigor of wheat under salinity stress. J. Stress Physiol. Biochem. 2005, 1, 6–14.
- 61. Angrish, A.; Kumar, B.; Datta, K.S. Effect of gibberellic acid and kinetin on nitrogen content and nitrate reductase activity in wheat under saline conditions. Indian J. Plant Physiol. 2001, *6*, 172-177.
- Gadallah, M.A.A. Effects of kinetin on growth, grain yield and some mineral elements in wheat plants growing under excess salinity and oxygen deficiency. Plant Growth Regul. 1999, 27, 63–74. doi: 10.1023/A:1006181204765.
- Datta, K.; Varma, S.; Angrish, R.; Kumar, B.; Kumari, P. Alleviation of salt stress by plant growth regulators in *Triticum aestivum* L. Biol. Plant. 1997, 40, 269-275. doi: 10.1023/A:1001076805595.
- Hedden, P.; Sponsel, V.A. Century of Gibberellin Research. J. Plant Growth Reg. 2015, 34, 740–760. doi: 10.1007/s00344-015-9546-1.
- Hasanuzzaman, M.; Bhuyan, M.H.; Parvin, K.; Bhuiyan, T.F.; Anee, T.I.; Nahar, K.; Hossen, M.; Zulfiqar, F.; Alam, M.; Fujita, M. Regulation of ROS metabolism in plants under environmental stress: A review of recent experimental evidence. Int. J. Mol. Sci. 2020, 22, 8695.
- Moumita, M.; Mahmud, J.A.; Biswas, P.K.; Nahar, K.; Fujita, M.; Hasanuzzaman, M. Exogenous application of gibberellic acid mitigates drought-induced damage in spring wheat. Acta Agrobot. 2019, 72, 1776. doi: 10.5586/aa.1776.
- 67. Jayasinghe, T.; Perera, P.; Wimalasekera, R. Effect of foliar application of gibberellin in mitigating salt stress in tomato (*Solanum Lycopersicum*), 'Thilina' variety; Proceedings of the 6<sup>th</sup> International Conference on Multidisciplinary Approaches [iCMA], Faculty of Graduate Studies, University of Sri Jayewardenepura; Nugegoda, Sri Lanka. 26–27 November 2019, [accessed on 3 December 2019]. Available online: https://ssrn.com/abstract=3497340.
- Miceli, A.; Moncada, A.; Sabatino, L.; Vetrano, F. Effect of gibberellic acid on growth, yield, and quality of leaf lettuce and rocket grown in a floating system. Agronomy. 2019, *9*, 382.
- Mittler, R.; Blumwald, E. The roles of ROS and ABA in systemic acquired acclimation. Plant Cell. 2015, 27, 64-70.
- 70. Devinar, G.; Llanes, A.; Masciarelli, O.; Luna, V. Different relative humidity conditions combined with

chloride and sulfate salinity treatments modify abscisic acid and salicylic acid levels in the halophyte Prosopis strombulifera. Plant Growth Reg. 2013, 70, 247-256.

- 71. Gurmani, A.R.; Bano, A.; Ullah, N.; Khan, H.; Jahangir, M.; Flowers, T.J. Exogenous abscisic acid (ABA) and silicon (Si) promote salinity tolerance by reducing sodium [Na+] transport and bypass flow in rice (*Oryza sativa indica*). Aust. J. Crop. Sci. 2013, 7, 1219-1226.
- 72. Liu, X.L.; Zhang, H.; Jin, Y.Y.; Wang, M.M.; Yang, H.Y.; Ma, H.Y.; Jiang, C.J.; Liang, Z.W. Abscisic acid primes rice seedlings for enhanced tolerance to alkaline stress by upregulating antioxidant defense and stress tolerance-related genes. Plant Soil. 2019, 26, 234-238.
- Bucker-Neto, L.; Paiva, A.L.S.; Machado, R.D.; Arenhart, R.A.; Margis-Pinheiro, M. Interactions between plant hormones and HMs responses. Genet. Mol. Biol. 2017, 40, 373-386.
- 74. Marthandan, V.; Geetha, R.; Kumutha, K.; Renganathan, V.G.; Karthikeyan, A.; Ramalingam, J. Seed Priming: A Feasible Strategy to Enhance Drought Tolerance in Crop Plants. Int. J. Mol. Sci. 2020, 21, 8258. doi: 10.3390/ijms21218258.
- 75. Miura, K.; Tada, Y. Regulation of water, salinity, and cold stress responses by salicylic acid. Frontiers in Plant Science. 2014, 5, 4.
- 76. Zhang, Y.; Xu, S.; Yang, S.; Chen, Y. Salicylic acid alleviates cadmium-induced inhibition of growth and photosynthesis through upregulating antioxidant defence system in two melon cultivars (*Cucumis melo* L.). Protoplasma. 2015, 252, 911–924.
- 77. Khan, M.I.R.; Asgher, M.; Khan, N.A. Alleviation of salt-induced photosynthesis and growth inhibition by salicylic acid involves glycinebetaine and ethylene in mungbean (*Vigna radiata* L.). Plant Physiol. Biochem. 2014, 80, 67-74.
- Fayez, K.A.; Bazaid, S.A. Improving drought and salinity tolerance in barley by application of salicylic acid and potassium nitrate. J. Saudi Soc. Agri. Sci. 2014, 13, 45-55.
- 79. Khan, M.I.R.; Iqbal, N.; Masood, A.; Per, T.S.; Khan, N.A. Salicylic acid alleviates adverse effects of heat stress on photosynthesis through changes in proline production and ethylene formation. Plant Signal. Behav. 2013b, 8, e26374.
- Habibi, A.; Abdoli, M. Influence of salicylic acid pretreatment on germination, vigour and growth parameters of garden cress (*Lepidium sativum*) seedlings under water potential loss at salinity stress. Int. Res. J. Basic Appl. Sci. 2013, 4, 1393-1399.
- Chavoushi, M.; Najafi, F.; Salimi, A.; Angaji, S.A. Improvement in drought stress tolerance of safflower during vegetative growth by exogenous application of salicylic acid and sodium nitroprusside. Ind. Crops Prod. 2019, 134, 168-176.

- Rehman, H.; Farooq, M.; Basra, S.M.; Afzal, I. Hormonal priming with salicylic acid improves the emergence and early seedling growth in cucumber. J. Agric. Soc. Sci. 2011, 7, 109–113.
- Soliman, M.H.; Al-Juhani, R.S.; Hashash, M.A.; Al-Juhani, F.M. Effect of seed priming with salicylic acid on seed germination and seedling growth of broad bean (*Vicia faba* L.). Int. J. Agric. Technol. 2016, 12, 1125-1138.
- Ahmad F., Iqbal S., Khan M.R., Abbas M.W., Ahmad J., Nawaz H., Shah S.M., Iqbal S., Ahmad M., Ali M. Influence of seed priming with salicylic acid on germination and early growth of sesame. Pure Appl. Biol. 2019, 8, 1206-1213. doi: 10.19045/bspab.2019.80062.
- 85. Shinwari, K.I.; Jan, M.; Shah, G.; Khattak, S.R.; Urehman, S.; Daud, M.K.; Naeem, R.; Jamil, M. Seed priming with salicylic acid induces tolerance against chromium toxicity in rice (*Oryza sativa* L.). Pak. J. Bot. 2015, 47, 161-170.
- Shatpathy, P.; Kar, M.; Dwibedi, S.K.; Dash, A. Seed priming with salicylic acid improves germination and seedling growth of rice (*Oryza sativa* L.) under PEG-6000 induced water stress. Int. J. Cur. Microbiol. Appl. Sci. 2018, 7, 907–924. doi: 10.20546/ijcmas.2018.710.101.
- Pouramir-Dashtmian, F.; Khajeh-Hosseini, M.; Esfahani, M. Improving chilling tolerance of rice seedling by seed priming with salicylic acid. Arch. Agron. Soil Sci. 2014, 60, 1291-1302. doi: 10.1080/03650340.2014.892584.
- Theerakulpisut, P.; Kanawapee, N.; Panwong, B. Seed priming alleviated salt stress effects on rice seedlings by improving Na<sup>+</sup>/K<sup>+</sup> and maintaining membrane integrity. Int. J. Plant Biol. 2016, 7, 6402. doi: 10.4081/pb.2016.6402.
- Chavoushi M.; Najafi, F.; Salimi, A.; Angaji, S.A. Improvement in drought stress tolerance of safflower during vegetative growth by exogenous application of salicylic acid and sodium nitroprusside. Ind. Crops Prod. 2019, 134, 168-176. doi: 10.1016/j.indcrop.2019.03.071.
- 90. Li, Z.; Xu, J.; Gao, Y.; Wang, C.; Guo, G.; Luo, Y.; Huang, Y.; Hu, W.; Sheteiwy, M.S.; Guan, Y. The synergistic priming effect of exogenous salicylic acid and H<sub>2</sub>O<sub>2</sub> on chilling tolerance enhancement during maize (*Zea mays* L.) seed germination. Front. Plant Sci. 2017, 8, 1153. doi: 10.3389/fpls.2017.01153.
- Farooq, M.; Aziz, T.; Basra, S.M.; Cheema, M.A.; Rehman, H. Chilling tolerance in hybrid maize induced by seed priming with salicylic acid. J. Agron. Crop. Sci. 2008, 194, 161-168. doi: 10.1111/j.1439-037X.2008.00300. x.
- 92. Zanganeh, R.; Jamei, R.; Rahmani, F. Impacts of seed priming with salicylic acid and sodium hydrosulfide on

possible metabolic pathway of two amino acids in maize plant under lead stress. Mol. Biol. Res. Commun. 2018, 7, 83-88. doi: 10.22099/mbrc.2018.29089.1317.

- 93. Singh, V.P.; Kumar, J.; Singh, M.; Singh, S.; Prasad, S.M.; Dwivedi, R.; Singh, M.P. Role of salicylic acid-seed priming in the regulation of chromium and UV-B toxicity in maize seedlings. Plant Growth Reg. 2016, 78, 79-91. doi: 10.1007/s10725-015-0076-4.
- 94. Afzal, I.; Basra, S.M.; Farooq, M.; Nawaz, A. Alleviation of salinity stress in spring wheat by hormonal priming with ABA, salicylic acid and ascorbic acid. Int. J. Agric. Biol. 2006, 8, 23-28.
- 95. Ulfat, A.N.; Majid, S.A.; Hameed, A. Hormonal seed priming improves wheat (*Triticum aestivum*) field performance under drought and non-stress conditions. Pak. J. Bot. 2017, 49, 1239-1253.
- Khamseh, S.R.; Shekari, F.; Zangani, E. The effects of priming with salicylic acid on resistance to osmotic stress in germination stage of wheat. Int. J. Agric. Res. Rev. 2013, 3, 543-558.
- 97. Azeem, M.; Abbasi, M.W.; Qasim, M.; Ali, H. Salicylic acid seed priming modulates some biochemical parameters to improve germination and seedling growth of salt stressed wheat (*Triticum aestivum* L.) Pak. J. Bot. 2018, 51, 385–391. doi: 10.30848/PJB2019-2[1].
- 98. El-Shazoly, R.M.; Metwally, A.A.; Hamada, A.M. Salicylic acid or thiamin increases tolerance to boron toxicity stress in wheat. J. Plant Nutr. 2019, 42, 702-722. doi: 10.1080/01904167.2018.1549670.
- 99. Gul, F.; Arfan, M.; Shahbaz, M.; Basra, S. Salicylic acid seed priming modulates morphology, nutrient relations and photosynthetic attributes of wheat grown under cadmium stress. Int. J. Agric. Biol. 2020, 23, 197-204. doi: 10.17957/IJAB/15.1277.
- 100. Namdari, A.; Baghbani, A. Consequences of seed priming with salicylic acid and hydro priming on smooth vetch seedling growth under water deficiency. J. Agric. Sci. 2017, 9, 259–267. doi: 10.5539/jas. v9n12p259.
- 101. Bahadoori, S.; Behrooz, E.; Mokhtar, H.; Surur, K.; Mosadegh, P.S.; Hassankeloo, N.T.; Alireza, G. Effects of seed priming with salicylic acid and polyamines on physiological and biochemical characteristics of okra (*Abelmoschus esculentus*) under low temperature stress. J. Plant Process Funct. 2016, *5*, 145–156.
- 102. Tabatabaei, S.A. Effect of salicylic acid and ascorbic acid on germination indexes and enzyme activity of sorghum seeds under drought stress. J. Stress Physiol. Biochem. 2013, 9, 32-38.
- 103. Ghoohestani, A.; Gheisary, H.; Zahedi, S.; Dolatkhahi, A. Effect of seed priming of tomato with salicylic acid, ascorbic acid and hydrogen peroxide on germination

and plantlet growth in saline conditions. Int. J. Agron. Plant Prod. 2012, 3, 700-704.

- 104. Singh, S.K.; Singh, P.K. Effect of seed priming of tomato with salicylic acid on growth, flowering, yield and fruit quality under high temperature stress conditions. Int. J. Adv. Res. 2016, 4, 723-727.
- 105. Rafique, N.; Raza, S.H.; Qasim, M.; Iqbal, N.A. Presowing application of ascorbic acid and salicylic acid to seed of pumpkin and seedling response to salt. Pak. J. Bot. 2011, 43, 2677–2682.
- 106. Azooz, M.M. Salt stress mitigation by seed priming with salicylic acid in two faba bean genotypes differing in salt tolerance. Int. J. Agric. Biol. 2009, 11, 343–350.
- 107. Khan, M.I.R.; Khan, N.A. Ethylene reverses photosynthetic inhibition by nickel and zinc in mustard through changes in PS II activity, photosynthetic nitrogen use efficiency, and antioxidant metabolism. Protoplasma. 2014, 251, 1007-1019.
- 108. Wang, K.L.; Li, H.; Ecker, J.R. Ethylene biosynthesis and signalling networks. Plant Cell. 2002, 14, S131–S151.
- 109. Khan, N.A.; Asgher, M.; Per, T.S.; Masood, A.; Fatma, M.; Khan, M.I.R. Ethylene Potentiates Sulfur-Mediated Reversal of Cadmium Inhibited Photosynthetic Responses in Mustard. Frontiers in Plant Science. 2016, 7.
- 110. Juvany, M.; Müller, M.; Munné-Bosch, S. Photooxidative stress in emerging and senescing leaves: a mirror image? J. Exp. Bot. 2013, 64(11), 3087–3098.
- 111. Jisha, K.C.; Vijayakumari, K.; Puthur, J.T. Seed priming for abiotic stress tolerance: an overview. Acta Physiol. Plant. 2013, 35, 1381–1396.
- 112. Wei, T.J.; Wang, M.M.; Jin, Y.Y.; Zhang, G.H.; Liu, M.;Yang, H.Y.; Jiang, C.J.; Liang, J.W. Abscisic acid priming creates alkaline tolerance in Alfalfa seedlings (*Medicago sativa* L.). Agriculture, 2021, 11, 608.

- 113. Hussain, S.; Zhu, C.; Huang, J.; Huang, J.; Zhu, L.; Cao, X.; Nanda, S.; Khaskheli, M.A.; Liang, Q.; Kong, Y.; Jin, Q.; Zhang, J. Ethylene response of salt stressed rice seedlings following Ethephon and 1-methylcyclopropene seed priming. Plant Growth Reg. 2020, 92, 219–231.
- 114. Wang C.P.; Chen, J.W.; Qiao, G.X. Proline inhibits plant root growth through signal of auxin pathway in Arabidopsis thaliana. *Zhiwu Shengli Xuebao*/Plant Physiol.<u>J.</u> 2017, 53(8), 1428-1434.
- 115. Gavelienė, V.; Pakalniškytė, L.; Novickienė, L. Regulation of proline and ethylene levels in rape seedlings for freezing tolerance. Cent. Eur. J. Biol. 2014, 9(11), 1099-1107.
- 116. Ábrahám E.; Rigó G.; Székely G.; Nagy R.; Koncz C.; Szabados L. Light-dependent induction of proline biosynthesis by abscisic acid and salt stress is inhibited by brassinosteroid in Arabidopsis. Plant Mol. Biol. 2003, 51, 363–372.
- 117. Sharma S.; Verslues P.E. Mechanisms independent of abscisic acid (ABA) or proline feedback have a predominant role in transcriptional regulation of proline metabolism during low water potential and stress recovery. Plant Cell Environ. 2010, 33, 1838–1851.
- 118. Liang, W.; Liang, L.; Ji, Z.; Li, P. The fluctuation of endogenous gibberellin and indole-3-acetic acid in *Litchi chinensis* shoot-tips during floral initiation. Acta Hort. Sinica. 1987; 14:145-152.