Journal of Agricultural, Food Science & Biotechnology

Short Communication

Flavoring profiles of Artemisia ludoviciana and Matricaria discoidea (Asteraceae) from Northeastern Utah

Katharina E. C. Wingard , Isabel P. Lykken , Tyler M. Wilson* and Christopher R.

D. Gary Young Research Institute, Lehi, UT, USA, 84043

Article Information

Received: 03 November 2025 Revised: 15 November 2025 Accepted: 20 November 2025 Published: 24 November 2025

Academic Editor

Prof. Dr. Gian Carlo Tenore

Corresponding Author

Prof. Dr. Tyler M. Wilson

tywilson@youngliving.com; Tel: Tel.: +1-801-669-4501

Keywords

Artemisia ludoviciana, ethanol extraction, flavor profile, Matricaria discoidea.

Abstract

Artemisia ludoviciana and Matricaria discoidea are aromatic plants in the Asteraceae family. Both plant species, which exhibit large native geographic ranges, have broad historical usage, including as food and flavoring materials. In the current study, flavoring profiles were established from populations originating in the Intermountain Region (USA), specifically in Northeastern Utah. Ethanol extractions (maceration technique) were analyzed by GC/MS. The flavor profile of A. ludoviciana was primarily composed of 1,8cineole (43.7%), camphene (18.4%), camphor (10.8%), α -pinene (6.8%), sabinene (6.7%), santolina triene (4.6%), and β -pinene (3.6%). The flavor profile of M. discoidea was simpler, with prominent compounds being myrcene (63.5%), (E)-β-farnesene (19.4%), limonene (11.4%), α -pinene (2.9%), and β -pinene (2.3%). The prominent volatile compounds present in A. ludoviciana are characterized by flavor profiles of camphor, earthy, eucalyptol, and spice flavors. Those abundant in M. discoidea are characterized as having flavor profiles of fruit, herb, citrus, and sweet flavors. The established aromatic profiles provide a foundation for the use and application of these plants in the food and flavor industry.

1. Introduction

Artemisia ludoviciana Nutt., also known as white sagebrush, is an aromatic plant in the Asteraceae family [1,2]. This perennial flowering plant is native to the Intermountain Region, east throughout the USA, south throughout Central America, and north into Canada [2-6]. A. ludoviciana has been used, typically as a tea or infusion, to help with a wide range of aliments including sores, respiratory issues, eczema, arthritis, horse illnesses, and has been used in many cleansing-spiritual rituals [1, 7-11]. However, throughout history, the most common use of the plant has been to treat infectious diseases and their symptoms [1, 5, 7-10]. Previous research of the leaf volatiles (flavoring compounds), determined by GC

analysis, of A. ludoviciana have been identified as being largely composed of β -ocimene, α -pinene, camphene, hexyl butyrate, 1,8-cineole, camphor, borneol, nonanal, linalool, carvacrol, and p- α dimethylbenzyl alcohol [8, 12].

Matricaria discoidea DC., also known as pineappleweed, is an annual plant of the Asteraceae family [3, 4, 13]. The exact origin of the species is debated, but it is believed to be native to both North America and Asia, and has been widely naturalized throughout Europe [14]. This flowering plant thrives in heavily treaded areas, such as roadsides and footpaths, and is characterized by its yellow, cone-shaped, rayless florets, feathery leaves, and distinct pineapple aroma



when crushed [14, 15]. M. discoidea, like its relative Roman chamomile (which is also often consumed as an infusion), is believed to have sedative properties, may help with anxiety and insomnia, has been noted for its anti-inflammatory properties, and has welldocumented ethnomedicinal uses of the entire plant [16]. Ouercetin galactoside, malonylapigenin glucoside, apigenin acetylglucoside, quercetin, luteolin, and apigenin glycosides are flavonoids that are relevant constituents of M. discoidea, and may be responsible for various purported pharmacological activities [17]. Volatile compounds (flavoring compounds) reportedly present in the essential oil of M. discoidea, as determined by GC analysis, include (Z)-enyne-dicycloether, (E)- β -farnesene, geranyl isovaleriate, and myrcene [18].

Given the widespread distribution and historical use of both plants in the Asteraceae family, chemical profiling of distinct regions and climates is of interest to the food and flavor. industries. The current study investigated the volatile profiles of ethanol extract of *A. ludoviciana* and *M. discoidea* from Northeastern Utah.

2. Materials and methods

2.1. Plant material collection

Artemisia ludoviciana and Matricaria discoidea (Fig. 1) populations grow naturally on public lands (Bureau of Land Management) in the Otter Creek area in Northeastern Utah (41°42′03.5″ N,111°22′.01.0″ W). Seeds were collected from this site during the fall of



Figure 1. Botanical illustration of *Artemisia ludoviciana* (left) and *Matricaria discoidea* (right). Illustration by Zach Nielsen.

2024, grown and cultivated in the Young Living Essential Oils (40°25'16.8"N greenhouse 111°52'24.3"W) during the winter of 2025, and transplanted to private garden plots in the Salt Lake Valley in the spring of 2025. Respecting the wishes of the landowner, the GPS coordinates for the private garden plots were withheld. Plant material (leaf, stem, and flower) for both species was harvested from the private garden plots on August 1, 2025 and allowed to shade-dry for 72 hours. A representative voucher sample of each species is held at the Young Living Aromatic Herbarium (YLAH): Artemisia ludoviciana Nutt., Wilson 2025-01, and Matricaria discoidea DC., Wilson 2025-01.

2.2. Extraction technique

Laboratory-scale extractions were performed using a custom 2-L stainless steel device (Fig. 2) as follows: 500 mL of 200-proof ethanol (MilliporeSigma, Sigma Aldrich, St. Louis, MS, USA) were combined with 100 g of plant material and macerated for 72 h. The plant material was then removed, and the liquid solution was heated and distilled for 10 min after passover. The condensed distillate was stored in a glass amber bottle until analysis.



Figure 2. Illustration of distillation unit employed. Section 1 (2-L capacity) holds ethanol and plant materials (maceration) for 72-hours, upon heating the vapor passes through section 2, and the vapor is cooled and condensed back to a liquid state in section 3. Illustration by Rick Simonson (Science Lab Studios, Inc).

2.3. Analysis methods

To determine volatile compound profiles, samples

Table 1. Volatile compounds detected in Artemisia ludoviciana and Matricaria discoidea ethanol extractions.

Compounds	KI	Relative area %	
		A. ludoviciana	M. discoidea
Santolina triene	906	4.6	nd
Tricyclene	921	0.9	nd
α -Thujene	924	0.3	nd
α -Pinene	932	6.8	2.9
Camphene	946	18.4	nd
Sabinene	969	6.7	nd
β-Pinene	974	3.6	2.3
Myrcene	988	nd	63.5
p-Cymene	1020	0.8	nd
Limonene	1024	0.7	11.4
1,8-Cineole	1026	43.7	nd
Santolina epoxide	*1054	0.6	nd
Artemisia alcohol	1080	0.6	nd
Camphor	1141	10.8	nd
Artemisyl acetate	1169	0.3	nd
Bornyl acetate	1284	0.2	nd
(E)- $β$ -Farnesene	1454	nd	19.4
	Total	98.9	99.5

The compound name, KI, and relative area % are reported. Values are reported as an average of three repeat injections from the same sample (σ < 1 for all compounds). KI is the Kovat's Index value and was previously calculated by Robert Adams using a linear calculation on a DB-5 column [19]. * KI manually calculated.

were analyzed, and compounds were identified and quantified by GC/MS using an Agilent 7890B GC/5977B MSD (Agilent Technologies, Santa Clara, CA, USA) and Agilent J&W DB-5, 60 m × 0.25 mm, 0.25 µm film thickness, fused silica capillary column. Operating conditions: 0.1 µL of sample (splitless injection) was injected using an autosampler, the initial oven temperature was set to 40 °C with an initial hold time of 5 min, and an oven ramp rate of 4.5 °C per min to 310 °C with a hold time of 5 min. The electron ionization energy was 70 eV, the scan range was 35-650 amu, the scan rate was 2.4 scans per second, the source temperature was set to 230 °C, and the quadrupole temperature was set to 150 °C. Compounds were identified using the Adams volatile oil library [19] and a Chemstation library (MSD Chemstation F.01.03.2357) search in conjunction with retention indices. Samples were analyzed in triplicate and the reported values (relative area %) are an average of those relative quantifications (σ < 1 for all compounds).

3. Results

Ethanol distillations (maceration technique) were

conducted on both Artemisia ludoviciana and Matricaria discoidea. The volatile profile of A. ludoviciana comprised compounds with prominent compounds (compounds > 2%) being 1,8-cineole (43.7%), camphene (18.4%), camphor (10.8%), α pinene (6.8%), sabinene (6.7%), santolina triene (4.6%), and β -pinene (3.6%). The volatile profile of M. discoidea was composed of the following five compounds: myrcene (63.5%), (E)- β -farnesene (19.4%), limonene (11.4%), α -pinene (2.9%), and β pinene (2.3%). The profiles are detailed in Table 1.

4. Discussion

Samples (n = 2) of *Artemisia ludoviciana* and *Matricaria discoidea* were produced by ethanol extraction. The exact technique (maceration technique) is often employed by distilleries for producing liquors such as gin. In this technique, aromatic herbs are soaked in an ethanol solution for a prolonged period depending on the structural and quality characteristics of the botanicals used. For this study, a 72-h maceration period was employed, which was suggested by industry experts [20]. The 72-h maceration technique was employed as a viable and representative

approach for recovering volatile compounds (flavoring compounds) that would be of interest to the food and flavor industry.

Previous research found that the volatile profile of *A*. ludoviciana is largely composed of β -ocimene, α pinene, camphene, hexyl butyrate, 1,8-cineole, camphor, borneol, nonanal, linalool, carvacrol, and p- α -dimethylbenzyl alcohol [8, 12]. In the current study, there was considerable overlap in the volatile profile of A. ludoviciana, with prominent compounds being 1,8-cineole (43.7%), camphene (18.4%), camphor (10.8%), α -pinene (6.8%), sabinene (6.7%), santolina triene (4.6%), and β -pinene (3.6%). Similarly, there is much overlap with previous research on the volatile profile of M. discoidea, where previous research (Z)-enyne-dicycloether, determined that farnesene, geranyl isovaleriate, and myrcene are prevalent compounds [18]. Two of these compounds, myrcene (63.5%) and (E)- β -farnesene (19.4%), were also the most prevalent compounds in the sample from the current study. Other prominent compounds included limonene (11.4%), α -pinene (2.9%), and β pinene (2.3%). For both plant species, aromatic profiles likely differ due to variations in extraction techniques, compared to those employed in the current study, and due to the inherent variations in plant species with large native growing regions.

The Flavor and Extract Manufacturers Association of the United States (FEMA) has detailed the flavor profiles associated with many volatile compounds [21]. The descriptive quality for the top three most abundant volatile compounds in A. ludoviciana (camphene, 1,8-cineole, camphor) are associated with camphor, earthy, eucalyptol, and spice flavors [22-24]. The descriptive quality for the top three most abundant volatile compounds in M. discoidea limonene, and (E)- β -farnesene) associated with fruit, herbs, citrus, and sweet flavors [25-27]. While consumer taste experiences are somewhat subjective, the authors believe that these are accurate descriptive flavor profiles for the ethanol extracts A. ludoviciana and M. discoidea.

In the United States, the Food and Drug Administration (FDA) is responsible for controlling the food ingredients permitted for use in the food and flavor industry, including alcoholic beverages [28]. The

Code of Federal Regulations recognizes some species of *Artemisia* (*A. dracunculus*) and *Matricaria* (*M. chamomilla*), as generally recognized as safe food items [29]. Additionally, thujone-free *Artemisia* species are permitted for use in food items by the FDA [30]. While neither plant species, *A. ludoviciana* and *M. discoidea*, are specifically detailed as being permitted for use as food items within the United States, both species have widespread historical use as food items, contain aromatic profiles comprised of common natural compounds, and did not contain (in the case of *A. ludoviciana*) detectable amounts of thujone. As such, volatile extracts from both plant species could be considered as usable food ingredients within the United States if approved by the FDA.

Given the process of the maceration technique employed herein, it is likely that non-volatile compounds are also extracted from the plant materials of both species. This was further confirmed by the fact that after the 72-h period, the samples were dark green (*A. ludoviciana*) and yellow-green (*M. discoidea*). However, upon distillation, both samples were a clear, colorless solution, suggesting that non-volatile compounds were not retained during the distillation process. Future research should focus on the identification and quantification of these non-volatile compounds.

5. Conclusions

The current study established the volatile compound profiles of two plants in the Asteraceae family, *Artemisia ludoviciana* and *Matricaria discoidea*, using the maceration technique. These plant species have a large native distribution range; however, the current study focused on a small population in Northeastern Utah (USA).

The volatile profiles established herein reflect the extraction techniques employed by distilleries for flavoring beverages, such as gin. The profile for *A. ludoviciana* was primarily composed of 1,8-cineole (43.7%), camphene (18.4%), camphor (10.8%), α -pinene (6.8%), sabinene (6.7%), santolina triene (4.6%), and β -pinene (3.6%). The profile of *M. discoidea* was primarily composed of myrcene (63.5%), (*E*)- β -farnesene (19.4%), limonene (11.4%), α -pinene (2.9%), and β -pinene (2.3%). Prominent volatile compounds

present in *A. ludoviciana* portray camphor, earthy, eucalyptol, and spice flavors. Those abundant in *M. discoidea* portray fruit, herb, citrus, and sweet flavors. These findings are of interest to the food and flavor industry, which is investigating the use of botanicals native to the Intermountain Region (USA).

Disclaimer (artificial intelligence)

Author(s) hereby state that no generative AI tools such as Large Language Models (ChatGPT, Copilot, etc.) and text-to-image generators were utilized in the preparation or editing of this manuscript.

Authors' contributions

Conceptualization, T.M.W.; sample procurement and production, K.E.C.W., I.P.L., T.M.W.; methodology, T.M.W; software, T.M.W.; validation, C.R.B.; formal analysis, T.M.W.; data curation, T.M.W.; writing—original draft preparation, K.E.C.W., I.P.L., T.M.W.; writing—review and editing, K.E.C.W., I.P.L., C.R.B.; funding acquisition, C.R.B. All authors have read and agreed to the published version of the manuscript.

Acknowledgements

The authors wish to thank the Bureau of Land Management for permitting plant research on public lands, Zach Nielsen for the botanical illustrations (*Artemisia ludoviciana, Matricaria discoidea*), and Rick Simonson (Science Lab Studios, Inc.) for the technical illustration of the distillation unit.

Funding

This research was funded by Young Living Essential Oils.

Availability of data and materials

All data have been presented within the current manuscript. Additional data will be made available on request according to the journal policy.

Conflicts of interest

The authors declare no conflicts of interest.

References

 Stevens, M.; Roberts, W. White Sage Artemisia ludoviciana Nutt. Available online: https:// plants.usda.gov/DocumentLibrary/plantguide/pdf/p

- g_arlu.pdf (accessed on 9 June 2025).
- The World Flora Online. Available online: http://www.worldfloraonline.org/taxon/wfo-0000003 410 (accessed on 29 May 2025).
- 3. Cronquist, A.; Holmgren, A.H.; Holmgren, N.H.; Reveal, J.L. Intermountain Flora–Vascular Plants of the Intermountain West, USA, Volume 1; The New York Botanical Garden: Bronx, NY, USA, pp.232-234, 1972.
- Welsh, S.L.; Atwood, N.D.; Goodrich, S.; Higgins, L.C. A Utah Flora, 5th Edition; Brigham Young University: Provo, UT, USA, pp.20-21, 2016.
- 5. Alenazi, M.F.; Sundberg, M.D. Phylogenetic tree of *Artemisia* species (*A. Campestris, A. Carruthii, A. Dracunculus, A. Filifolia* and *A. Ludoviciana*) in Kansas based on morphological and anatomical characteristics. Trans. Kans. Acad. Sci. 2019, 122 (1/2), 29–38. http://www.jstor.org/stable/45106607
- Estes, J.R. Evidence for autoploid evolution in the Artemisia ludoviciana complex of the Pacific Northwest. Brittonia. 1969, 21, 29–43. https://doi.org/10.2307/2805657
- 7. Swor, K.; Poudel, A.; Satyal, P.; Setzer, W.N. The essential oil compositions of *Ambrosia acanthicarpa* Hook., *Artemisia ludoviciana* Nutt., and *Gutierrezia sarothrae* (Pursh) Britton & Rusby (Asteraceae) from the Owyhee Mountains of Idaho. Molecules, 2024, 29(6), 1383. https://doi.org/10.3390/molecules29061383
- 8. Lopes-Lutz, D.; Alviano, D.S.; Alviano, C.S.; Kolodziejczyk, P.P. Screening of chemical composition, antimicrobial and antioxidant activities of *Artemisia* essential oils. Phytochem. 2008, 69(8), 1732-1738.
 - https://doi.org/10.1016/j.phytochem.2008.02.014
- Romero, J.L.G.; Sosa, C.M.P.; Burgoa, G.L.; Leal, A.C.L.; El Kassis, E.G.; Rodríguez, E.B.; Juárez, G.A.P.; Hernández, L.R.; Bach, H.; Juárez, Z.N. Antimycobacterial, cytotoxic, and anti-inflammatory activities of *Artemisia ludoviciana*. J. Ethnopharmacol. 2022, 293, 115249.
 - https://doi.org/10.1016/j.jep.2022.115249
- McCutcheon, A.R. Ethnopharmacology of western North American plants with special focus on the genus *Artemisia L.* (T). Available online: https://open.library.ubc.ca/collections/ubctheses/831/i tems/1.0087277 (accessed 9 June 2025).
- Lans, C. Possible similarities between the folk medicine historically used by first nations and American Indians in North America and the ethnoveterinary knowledge currently used in British Columbia, Canada. J. Ethnopharmacol. 2016, 192, 53-66. https://doi.org/10.1016/j.jep.2016.07.004

- 12. Korablova, O.; Levchuk, I.; Rakhmetov, D.; Palamar, V.; Sydor, B.; Shanaida, M. Comparative analysis of essential oils from three species of the genus *Artemisia* cultivated in Ukraine. Botanica Lithuanica. 2023, 29(2). https://doi.org/10.35513/Botlit.2023.2.1
- 13. The World Flora Online. Available online. http://www.worldfloraonline.org/taxon/wfo-000002 8105 (accessed on 29 May 2025).
- 14. Bioimages Virtual Field Guide (UK). Available online: https://www.bioimages.org.uk/html/Matricaria_disco idea.htm (accessed on 29 May 2025).
- Sepp, J.; Koshovyi, O.; Jakštas, V.; Žvikas, V.; Botsula, I.; Kireyev, I.; Severina, H.; Kukhtenko, O.; Põhako-Palu, K.; Kogermann, K. Phytochemical, pharmacological, and molecular docking study of dry extracts of *Matricaria discoidea* DC. with analgesic and soporific activities. Biomolecules. 2024, 14(3), 361. https://doi.org/10.3390/biom14030361
- Sah, A.; Naseef, P.P.; Kuruniyan, M.S.; Jain, G.K.;
 Zakir, F.; Aggarwal, G. A Comprehensive study of therapeutic applications of chamomile.
 Pharmaceuticals. 2022, 15(10), 1284.
 https://doi.org/10.3390/ph15101284
- Koshovyi, O.; Sepp, J.; Jakštas, V.; Žvikas, V.; Tolmacheva, K.; Kireyev, I.; Heinämäki, J.; Raal, A. Phytochemical and pharmacological study on the dry extract of *Matricaria discoidea* DC. herb and its amino acids preparations. Phyton-Int. J. Exp. Bot. 2024, 93(11), 2909-2925. https://doi.org/10.32604/phyton.2024.056536
- Lopes, D.; Kolodziejczyk, P.P. Essential oil composition of pineapple-weed (*Matricaria discoidea* DC.) grown in canada. J. Essent. Oil-Bear. Plants. 2005, 8(2), 178–182. https://doi.org/10.1080/0972060X.2005.10643440
- R.P. Adams, Identification of Essential Oil Components by Gas Chromatography/Mass Spectrometry, 4th ed. Carol Stream, Allured Publ. (2007).
- 20. According to C. Barlow (Beehive Distilling) (personal communication 24 September 2025).

- The Flavor and Extract Manufacturers Association of the United States. Available online: https://www.femaflavor.org (accessed on 10 Nov 2025).
- The Flavor and Extract Manufacturers Association of the United States. Available online: https://www. femaflavor.org/flavor-library/camphene (accessed on 10 Nov 2025).
- The Flavor and Extract Manufacturers Association of the United States. Available online: https://www. femaflavor.org/flavor-library/eucalyptol (accessed on 10 Nov 2025).
- 24. The Flavor and Extract Manufacturers Association of the United States. Available online: https://www.femaflavor.org/flavor-library/d-camphor (accessed on 10 Nov 2025).
- The Flavor and Extract Manufacturers Association of the United States. Available online: https://www. femaflavor.org/flavor-library/myrcene(accessed on 10 Nov 2025).
- The Flavor and Extract Manufacturers Association of the United States. Available online: https://www. femaflavor.org/flavor-library/d-limonene(accessed on 10 Nov 2025).
- 27. The Flavor and Extract Manufacturers Association of the United States. Available online: https://www.femaflavor.org/flavor-library/beta-farnesene (accessed on 10 Nov 2025).
- 28. The Alcohol and Tobacco Tax and Trade Bureau. Available online: https://www.ttb.gov/scientific-services-division/limited-ingredients (accessed on 3 Nov 2025).
- 29. The Electronic Code of Federal Regulations, part 182. Available online: https://www.ecfr.gov/current/title-21/chapter-I/subchapter-B/part-182 (accessed on 3 Nov 2025).
- 30. The Electronic Code of Federal Regulations, part 172. Available online: https://www.ecfr.gov/current/title-21/chapter-I/subchapter-B/part-172 (accessed on 3 Nov 2025).